



## mTOR and cancer: many loops in one pathway

Alejo Efeyan<sup>1,2</sup> and David M Sabatini<sup>1,2,3</sup>

The mammalian target of rapamycin (mTOR) is a master regulator of cell growth and division that responds to a variety of stimuli, including nutrient, energy, and growth factors. In the last years, a significant number of pieces have been added to the puzzle of how mTOR coordinates and executes its functions. Extensive research on mTOR has also uncovered a complex network of regulatory loops that impact the therapeutic approaches aimed at targeting mTOR.

#### Addresses

- <sup>1</sup> Whitehead Institute for Biomedical Research, Nine Cambridge Center, Cambridge, MA 02142, United States
- <sup>2</sup> Howard Hughes Medical Institute, Department of Biology, Massachusetts Institute of Technology, Cambridge, MA 02139, United States
- <sup>3</sup> Koch Institute for Integrative Research in Cancer at MIT, 77 Massachusetts Avenue, Cambridge, MA 02139, United States

Corresponding author: Sabatini, David M (sabatini@wi.mit.edu)

#### Current Opinion in Cell Biology 2010, 22:169-176

This review comes from a themed issue on Cell regulation Edited by Guido Kroemer and Eileen White

Available online 27th November 2009

0955-0674/\$ - see front matter
© 2009 Elsevier Ltd. All rights reserved.

DOI 10.1016/j.ceb.2009.10.007

#### Introduction

Adequate cellular levels of energy and nutrients are a prerequisite for successful cell growth and division. Therefore, cells have acquired mechanisms to sense energy and nutrients before committing to grow and divide. In all eukaryotes, the target of rapamycin (TOR) protein is a master regulator that integrates the signals from nutrient and energy sensors with cell growth and proliferation, so as to ensure that they are triggered only during favorable conditions. The mammalian TOR (mTOR) also integrates growth factor signaling together with nutrients and energy as a mechanism to coordinate cell growth and proliferation in large number of cells within different organs. In addition to this, a network of regulatory loops affects the function of mTOR and impact on therapeutic approaches aimed at targeting mTOR, which will be discussed here.

#### The mTOR signaling pathway

mTOR forms two different protein complexes defined by the proteins to which it is bound, exerting different but related functions. The mTOR complex 1 (mTORC1) is defined by the presence of Raptor, mLST8/G $\beta$ L, Deptor,

and PRAS40 [1–4,5°,6,7], whereas Rictor, GβL, Protor, Deptor, and mSin form mTORC2 [8–12] (see Figure 1). In addition to their differential sensitivity to rapamycin, mTORC1 and 2 are activated in different ways and have distinct substrate specificity. mTORC1, which is sensitive to rapamycin, responds to energy, amino acids, growth factors, and oxygen levels, whereas mTORC2 activation is ill-defined, but seems to be mediated only by growth factors. Active mTORC1 phosphorylates, among other targets, S6K1 and 4EBP1, proteins involved in the regulation of translation initiation, protein synthesis, and cell mass gain (see [13°] for a comprehensive review). An overview of the mTOR complexes and signaling pathway is provided in Figure 1.

#### The Rag - amino acid link

The fact that amino acid-dependent activation of mTORC1 occurs independently of the Akt-TSC axis signaling has been known for years, but the identification of the molecular link between amino acids and mTORC1 has remained elusive. Recently, we and others showed that the Rag family of GTPases is required for mTORC1 activation by amino acids [14\*\*,15\*\*]. Rag proteins directly interact with Raptor in an amino acid-dependent manner, and this interaction leads to mTORC1 activation. Interestingly, the nucleotide-bound status of the Rag GTPases dictates its affinity to Raptor and the activation of mTORC1. This discovery shows a previously unknown regulatory mechanism of mTOR function that involves mTOR shuttling. In the presence of amino acids, Rag proteins mediate mTORC1 shuttle from disperse punctuate pattern in the cytoplasm to the endomembrane system of the cell, where signaling occurs [15\*\*]. According to this, appropriate localization of mTORC1 and active PI3K-Akt-Rheb axis must coexist, explaining why both amino acids and growth factors are needed for the activation of mTORC1. It is likely that Rag GTPases are not responsible for directly sensing amino acid levels; instead this information is probably communicated to the Rag proteins by additional mediators.

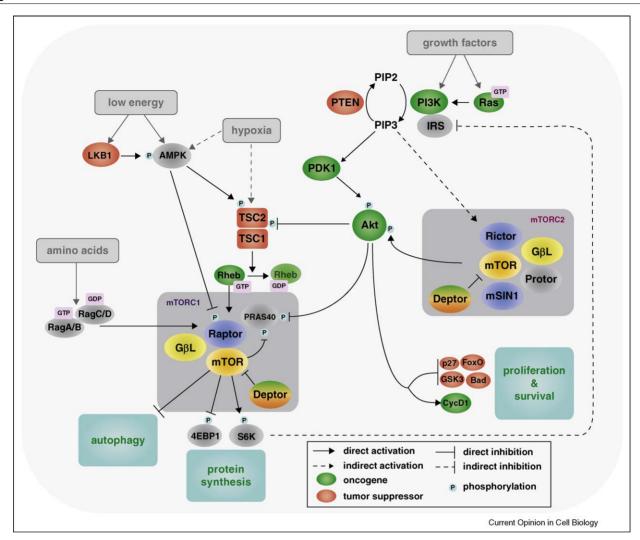
# Complex loops of regulation — all roads lead to mTOR

mTOR, as a master regulator of a variety of inputs, is subjected to different mechanisms that tightly and coordinately regulate its activation. Many positive and feedback loops have been described lately and most likely other regulatory loops will be deciphered in the near future.

#### The S6K1-PI3K inhibition

A very important negative feedback involves the inhibition of the PI3K pathway by mTORC1, which occurs at

Figure 1



mTOR signaling pathway. One branch of mTORC1 activation is mediated by the class I phosphoinositide-3-kinase (Pl3K), Akt (also known as Pkb) and the tuberous sclerosis complex (TSC). TSC is formed by TSC1 and TSC2, and inhibits a direct activator of mTORC1, the GTPase Ras-homolog enriched in brain (Rheb) by hydrolyzing its GTP into GDP [52]. TSC2 is activated by phosphorylation by AMP-activated protein kinase (AMPK) [53], which is directly activated by a high AMP versus ATP ratio. AMPK also directly phosphorylates and inactivates Raptor, so it inhibits mTORC1 by TSCdependent and TSC-independent manners [54]. The activity of AMPK is regulated by phosphorylation by the tumor suppressor LKB1. This protein, like TSC1/2, was found mutated in the germline of patients with different hamartomatous syndromes [55,56]. Akt is a serine/threonine kinase and an important player in regulating mTORC1 activity. Akt positively regulates mTORC1 by acting at different levels. First, Akt inactivates TSC1/2 by phosphorylating TSC2 [57]. Second, Akt inhibits PRAS40, negative regulator of mTORC1 that counteracts Rheb function [6,7]. Akt is activated by PI3K, which responds to a variety of growth factors. When activated by insulin or insulin-like growth factors (IGFs), as well as other growth factors, class I PI3K catalyzes the formation of the lipidic second messenger phosphoinositide-3,4,5-tri-phosphate (PIP3) from the bi-phosphate form PIP2. PIP3 triggers the relocation of Akt to the inner surface of the plasma membrane, where it is activated by phosphoinositide-dependent kinase 1 (PDK1) and transduces the signal as described above. Opposing Akt function is the tumor suppressor phosphatase and tensin homolog deleted on chromosome ten (PTEN), a lipid phosphatase that converts PIP3 to PIP2, thus shutting off signaling from PI3K. PTEN deficiency causes a series of hamartomatous syndromes collectively classified as PTEN hamartoma tumor syndrome (reviewed in [58]). Amino acids activate mTORC1 by an independent route mediated by the Rag family of proteins. The activation of mTORC2 is not well understood, but this complex directly activates Akt (and Akt-related kinases) by phosphorylation. Akt, in addition, regulates many proteins involved in cell survival and cell-cycle progression.

many levels (see Figure 2a). Active S6K inhibits insulin receptor substrate 1 (IRS-1) by phosphorylating it at multiple sites, by inducing its degradation and by altering its localization, all of which ultimately dampen PI3K/Akt

activation [16-19]. This loop is relevant in vivo and influences therapeutic responses based on mTOR inhibition (see below), and also plays a role in type 2 diabetes [20]. A functionally similar loop mediated by S6K1 is the

inhibition of platelet-derived growth factor receptor (PDGFR), that also signals through PI3K [21]. Of note, the direct target of S6K1 that inhibits PDGFR activity is currently unknown. In addition, it has been recently shown that S6K1 is also responsible for inhibiting the activity of the ERK/MAPK pathway. This loop was empirically unmasked by therapeutic inhibition of mTORC1 in cancer patients, which correlated with an increase in ERK phosphorylation in vivo [22]. In spite of the fact that most research on mTORC1 is centered on S6K1, it is also conceivable that the effects on mTORC1 inhibition may not be solely through this target. These partially overlapping inhibitory loops have probably evolved as a mechanism to negatively control PI3K pathway after its activation, reflecting the necessity of putting a brake to excessive (acute or chronic) activation of the pathway.

#### p53 and mTORC1

The tumor suppressor p53 is activated by different types of cellular stress, including DNA damage and oncogene activation. p53 mediates transactivation of a large number of genes involved in cell-cycle arrest and apoptosis. p53 also transactivates negative regulators of mTORC1, namely TSC2 and AMPKB1 [23]. Besides, AMPK phosphorylates p53 at the serine in position 15, which leads to stabilization and favors the transactivation capacity of p53 [24]. Hence, p53 and AMPK positively regulate each other. Moreover, p53 upregulates the transcription of Sestrins 1 and 2, which bind to the ternary complex TSC1/2-AMPK, inducing phosphorylation and activation of TSC2 by AMPK [25]. In addition PTEN was shown to interact with p53, leading to p53 stabilization [26]. Interestingly, Akt is a negative regulator of p53 activity by phosphorylating an E3 ubiquitin ligase, namely MDM2, which drives its translocation to the nucleus where it destabilizes p53 [27]. These cross regulations of AMPK, PTEN, and p53 oppose Akt activity (see Figure 2b) and constitute a network coordination of division and growth after sensing nutrient availability, DNA integrity, and oncogenic activity.

## Intra-complex loops

MTORC1 and 2 activities are affected by cross-regulatory loops triggered by components of both complexes. Perhaps the most interesting loop where both mTOR complexes participate is the one involving mTOR regulation of Akt. It has been shown that mTORC2 is the kinase of Akt at the serine in position 473 [28]. This phosphorylation, together with the one at 308 performed by PDK1 upon PI3K activation, is needed for maximal Akt activity and conceptually locates mTOR both upstream and downstream Akt (Figure 2c).

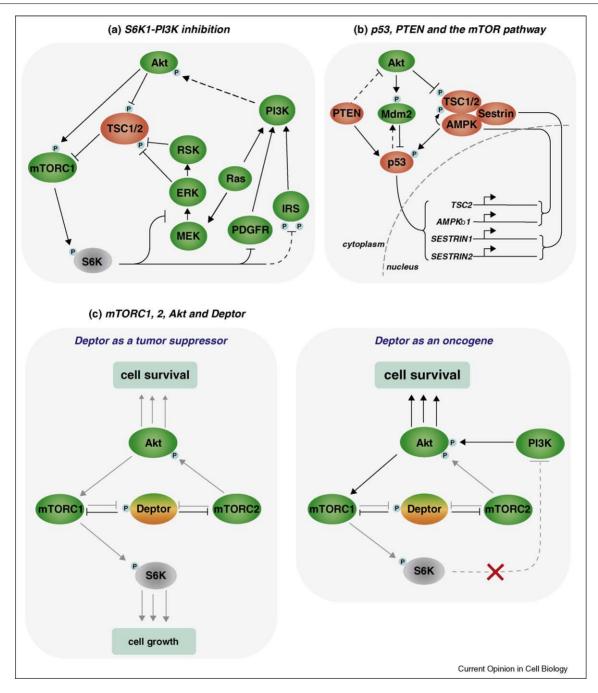
Our group has recently described the role of Deptor in the regulation of both mTORC1 and 2 [5°]. The functional relationship of Deptor and the mTORCs is not simple (Figure 2c). Deptor protein and mRNA levels are suppressed by mTORC1 and mTORC2 activation, and Deptor inhibits mTORC1 and 2 kinase activities in vitro and in cell-based assays. Consequently, the activation of mTORC1 and 2 leads to Deptor inhibition, leading to further mTOR activity and in contrast, high levels of Deptor inhibit mTOR. Importantly, the inhibition of mTORC1 by Deptor indirectly activates Akt, by relieving the inhibitory loop on PI3K triggered by S6K1. Considering this, theoretically Deptor could have both tumor suppressive (by the inhibition of mTORC1 and 2) and oncogenic roles (by the activation of Akt) (Figure 2c). Although more work and *in vivo* data will help clarify this issue, transcriptional profiling of tumor cell lines showed that Deptor was overexpressed in multiple myeloma cell lines that show high Akt activity. In these cell lines, silencing of Deptor leads to the inhibition of Akt and cell death, altogether suggesting that deregulated Deptor function may have oncogenic properties [5°].

Recently, the tuberous sclerosis complex (TSC), in addition to its role in the inhibition of mTORC1, was linked to mTORC2 function. TSC1/2 complex interacts with mTORC2 and positively regulates its kinase activity by an as-yet undefined mechanism that seems to be independent of the inhibition of mTORC1 [29°]. Although this finding requires further investigation, it locates TSC downstream and upstream of Akt. It also helps to explain the benign nature of tuberous sclerosis syndromes, because in TSC-deficient cells, in addition to the S6K1-PI3K-Akt feedback that inhibits Akt, its activity would be further diminished by reduced activation of mTORC2. In addition, S6K1 was shown to regulate mTORC2 activity by direct phosphorylation of the mTORC2 component Rictor. This phosphorylation event does not regulate mTORC2 kinase activity in vitro nor alters mTORC2 effects on most of its targets in cellbased experiments, but it negatively regulates phosphorylation of Akt at serine 473 [30]. These two new regulatory loops point toward a cellular mechanism that prevents simultaneous activation of mTORC1 and mTORC2, in addition to the already described role of S6K1 in putting a brake on excessive PI3K activation.

## The impact of mTOR feedback loops in cancer Rapamycin-based therapy

Extensive basic research with rapamycin showed high specificity toward mTORC1 inhibition. This has encouraged a number of clinical trials using this compound as an anticancer drug, but in fact, rapamycin and its first generation analogs temsirolimus and everolimus have proven modest success in clinical trials, reflecting an incomplete understanding of mTOR functions. These small molecules are allosteric inhibitors of mTOR that block S6K1 phosphorylation, but it has become evident that not all mTORC1 functions are blocked by rapamycin [31,32°,33°,34°,35°]. Rapamycin also stops the negative

Figure 2



Feedback loops modulating mTOR signaling pathway. (a) S6K1 phosphorylation by mTORC1 triggers a number of feedback loops that negatively impact on PI3K signaling. S6K1 directly phosphorylates insulin receptor substrate 1 (IRS-1) at multiple sites, which leads a significant attenuation of insulin and insulin-like growth factor effect at the cell membrane. S6K1 also inhibits PDGFR and the MEK/ERK signaling pathway through a not fully understood mechanism. These three inhibitory loops ultimately dampen PI3K, Akt, and mTORC1 activation and have likely evolved as a cellular response to buffer aberrant or excessive PI3K activity. (b) The two most important tumor suppressors, namely PTEN and p53, cooperate in the inhibition of mTORC1. p53 transactivates many negative regulators of mTORC1 (TSC2, AMPKβ1, Sestrins 1 and 2), which oppose Akt activity. Akt favors degradation of p53 by phosphorylating and stabilizing MDM2. Importantly, PTEN and AMPK directly stabilize p53. Collectively, these interactions imply the existence of a coordinate regulation of mTOR signaling pathway by energy, nutrients, growth factors, oncogenic stress, and DNA integrity. (c) Deptor can have oncogenic and tumor suppressive properties. By blocking mTORC1 and mTORC2, Deptor inhibits protein synthesis, cell size increase, and the proliferative and survival effects of Akt. However, under certain conditions, inhibition of mTORC1 by Deptor relieves the feedback inhibition from S6K1 to PI3K, boosting Akt activity.

feedback loops emanating from S6K1 to PI3K signaling pathway, thus activating proliferative and prosurvival effectors, as Akt. This was shown to be relevant in many cancer cell lines and also in clinical samples [36], and may explain why rapamycin, although slowing proliferation in many cell lines, is a poor inducer of apoptosis. Altogether, this raises an important concern: if mTORC1 has rapamycin-resistant functions and rapamycin indirectly activates the PI3K signaling pathway, which is responsible for mTORC1 activation, then rapamycin could cause hyperactivation of those mTORC1-dependent functions not inhibited by rapamycin.

Akt is not the only target downstream PI3K activation but there is in vivo evidence that points toward Akt as perhaps the most important target affected by the feedback loop from S6K1, and contributing to the modest therapeutic success of rapamycin. TSC2-heterozygous mice have constitutive mTORC1 activity and, consequently, overactivation of the feedback loop that dampens Akt activity. Interestingly, double heterozygous mice for TSC2 and PTEN have active Akt and accelerated tumorigenesis in some organs in comparison with single heterozygous mice [37,38]. Hay and colleagues [39] have shown that mTORC1 function downstream of PI3K-Akt is necessary for transformation and tumorigenesis in an experimental model of mammary and salivary tumors, favoring the hypothesis that rapamycin may be a good therapeutic strategy under conditions of Akt hyperactivation. However, Akt may drive tumorigenesis by regulating effectors different from mTORC1. In fact, Akt can regulate the Foxo proteins, cyclin D1, p27, and GSK3, all involved in cell-cycle progression, and MDM2, caspase-9, IKKa and Bad, controlling the execution of apoptosis [27]. Only in those tumors where mTORC1 is the important downstream effector of the oncogenic activity of Akt, rapamycin would be a reasonable therapeutic strategy.

PI3K also has Akt-independent effectors, with potential clinical relevance, that would also be indirectly activated by rapamycin. For instance, high levels of PDK1 can sustain tumor growth in an Akt-independent manner, by hyperactivation of SGK3 [40°]. SGK3 is part of the AGC family of proteins, that also includes Akt, and that shares target specificity. Interestingly, SGK proteins are, as Akt, substrates of mTORC2 [41].

Although rapamycin is a highly selective mTORC1 inhibitor, this has been challenged by the fact that it can also block mTORC2 activity in a subset of cancer cell lines after prolonged treatment through a mechanism that may involve inhibition of mTORC2 assembly [42]. Moreover, high concentration of rapamycin was also shown to block mTORC2 activity [43]. However, whether such toxic high levels of rapamycin have a clinical applicability is unclear.

#### mTOR catalytic inhibitors

The development of a different class of mTOR inhibitors that blocks mTOR catalytic site (Torin [34°], PP242 and PP30 [32°], Ku-0063794 [33°], and WAY-600, WYE-687, and WYE-354 [35°]) was reported almost simultaneously by several groups. This approach presents an obvious advantage for therapeutics: by blocking mTOR, both mTORC1 and mTORC2 become inhibited. Consequently, all mTOR target activities are blocked, inposition cluding Akt phosphorylation at [32°,33°,34°,35°]. The fact that mTORC2 is the kinase for Akt at Ser473 is now widely accepted and supported by genetic studies in mice. However, it is important to determine to what extent the inhibition of Ser473 phosphorylation impairs Akt activity toward all of its targets, which may be differentially affected [9,44]. How phosphorylation at positions 308 and 473 crossregulate Akt activity is particularly important in situations where S6K1 inhibition will boost PI3K activation (and, consequently, Akt phosphorylation at Thr308). Noteworthy, acute inhibition of mTORC2 using these mTOR catalytic inhibitors suggests that phosphorylation at Ser473 is required for phosphorylation at Thr308 and, consequently, for Akt activation. Further research should address how these compensatory loops would affect tumor responsiveness.

Toxicity is a possible caveat to consider if all mTOR functions are inhibited. However, work in mice suggests that blocking mTORC2 could be tolerated in vivo. In a murine model of PTEN deficiency-driven prostate tumorigenesis, deletion of Rictor (i.e. mTORC2) partially impairs prostate adenocarcinoma development, but does not affect normal prostate gland function and architecture [45°]. This indicates that mTORC2-specific inhibitors would be a good therapeutic strategy to be further investigated. In a similar experimental approach, Pandolfi and colleagues also showed that mTORC2 enhances prostate tumorigenesis, but ablation of mTOR in adult prostate does not affect normal prostate function [46°], providing strong evidence that mTOR catalytic inhibitors could be well tolerated in postmitotic cells. Supporting this notion, WYE-354 inhibited human cancer cells growing as xenografts in nude mice with no detectable toxicity in the short term [35°].

### **Dual PI3K-mTOR inhibitors**

An alternative way to circumvent the activation of feedback loops that may preclude tumor responsiveness is the combined inhibition of PI3K together with mTOR. There are several small molecules that block PI3K and mTORC1 and 2, currently under intense investigation (reviewed in [47]). In cell lines and experimental tumors with hyperactive PI3K signaling, dual inhibition of PI3K and mTOR was effective [48], but failed in the presence of K-Ras hyperactivation. In Ras-driven tumorigenesis, concomitant inhibition of PI3K and mTOR, and also blocking downstream mediators of Ras seem to be required [49–51]. Whether such a combinatorial signaling perturbation will be toxic also for normal cells is a reasonable caveat to be considered.

#### **Conclusions**

In the last years, substantial progression in the understanding of how mTOR signaling pathway occurs has taught us that there are still significant aspects of mTOR regulation that need further clarification. The mechanism underlying the induction of cell death versus cell-cycle arrest in different cell lines treated with mTOR inhibitors, the existence of additional feedback loops triggered by S6K1 and how these loops affect therapeutic responses should be addressed in the future. Moreover, a clearer picture of the relevance of Akt-dependent versus Akt-independent signaling of PI3K, how does phosphorylation of Akt at Thr308 and at Ser473 influence each other and Akt activity is also needed. Gaining insight into these issues will certainly settle the bases for better therapeutic strategies against cancer.

## **Acknowledgements**

We thank NIH and HHMI for funding our work on mTOR. A.E. is supported by the Human Frontiers Science Program.

#### References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- •• of outstanding interest
- Hara K, Maruki Y, Long X, Yoshino K, Oshiro N, Hidayat S, Tokunaga C, Avruch J, Yonezawa K: Raptor, a binding partner of target of rapamycin (TOR), mediates TOR action. Cell 2002, 110:177-189.
- Kim DH, Sarbassov DD, Ali SM, King JE, Latek RR, Erdjument-Bromage H, Tempst P, Sabatini DM: mTOR interacts with raptor to form a nutrient-sensitive complex that signals to the cell growth machinery. Cell 2002, 110:163-175.
- Kim DH, Sarbassov DD, Ali SM, Latek RR, Guntur KV, Erdjument-Bromage H, Tempst P, Sabatini DM: GbetaL, a positive regulator of the rapamycin-sensitive pathway required for the nutrientsensitive interaction between raptor and mTOR. Mol Cell 2003, 11:895-904.
- Loewith R, Jacinto E, Wullschleger S, Lorberg A, Crespo JL, Bonenfant D, Oppliger W, Jenoe P, Hall MN: Two TOR complexes, only one of which is rapamycin sensitive, have distinct roles in cell growth control. Mol Cell 2002, 10:457-468.
- Peterson TR, Laplante M, Thoreen CC, Sancak Y, Kang SA,
   Kuehl WM, Gray NS, Sabatini DM: DEPTOR is an mTOR inhibitor frequently overexpressed in multiple myeloma cells and required for their survival. Cell 2009, 137:873-886.

In this report, Deptor, a negative regulator of mTORC1 and 2, is identified. mTOR and Deptor inhibit each other, and interestingly, Deptor over-expression indirectly leads to Akt hyperactivation.

- Sancak Y, Thoreen CC, Peterson TR, Lindquist RA, Kang SA, Spooner E, Carr SA, Sabatini DM: PRAS40 is an insulinregulated inhibitor of the mTORC1 protein kinase. Mol Cell 2007, 25:903-915.
- Vander Haar E, Lee SI, Bandhakavi S, Griffin TJ, Kim DH: Insulin signalling to mTOR mediated by the Akt/PKB substrate PRAS40. Nat Cell Biol 2007, 9:316-323.
- 8. Frias MA, Thoreen CC, Jaffe JD, Schroder W, Sculley T, Carr SA, Sabatini DM: mSin1 is necessary for Akt/PKB phosphorylation,

- and its isoforms define three distinct mTORC2s. Curr Biol 2006, 16:1865-1870.
- Jacinto E, Facchinetti V, Liu D, Soto N, Wei S, Jung SY, Huang Q, Qin J, Su B: SIN1/MIP1 maintains rictor-mTOR complex integrity and regulates Akt phosphorylation and substrate specificity. Cell 2006, 127:125-137.
- Jacinto E, Loewith R, Schmidt A, Lin S, Ruegg MA, Hall A, Hall MN: Mammalian TOR complex 2 controls the actin cytoskeleton and is rapamycin insensitive. Nat Cell Biol 2004, 6:1122-1128.
- Sarbassov DD, Ali SM, Kim DH, Guertin DA, Latek RR, Erdjument-Bromage H, Tempst P, Sabatini DM: Rictor, a novel binding partner of mTOR, defines a rapamycin-insensitive and raptorindependent pathway that regulates the cytoskeleton. Curr Biol 2004, 14:1296-1302.
- Yang Q, Inoki K, Ikenoue T, Guan KL: Identification of Sin1 as an essential TORC2 component required for complex formation and kinase activity. Genes Dev 2006, 20:2820-2832.
- Ma XM, Blenis J: Molecular mechanisms of mTOR-mediated
   translational control. Nat Rev Mol Cell Biol 2009, 10:307-318.
   Comprehensive review of mTORC1 function, particularly in the regulation of translational control.
- Kim E, Goraksha-Hicks P, Li L, Neufeld TP, Guan KL: Regulation of TORC1 by Rag GTPases in nutrient response. Nat Cell Biol 2008, 10:935-945.

See annotation to Ref. [15\*\*].

Sancak Y, Peterson TR, Shaul YD, Lindquist RA, Thoreen CC, Bareled L, Sabatini DM: The Rag GTPases bind raptor and pate of the control of the contr

mediate amino acid signaling to mTORC1. Science 2008, 320:1496-1501.

Along with Ref. [14\*\*], this study identifies the Rag proteins as the direct link of amino acid-dependent activation of mTORC1. This study also demonstrates direct binding of Rag protein to mTORC1 and Rag-dependent mTORC1 shuttling as a novel regulatory mechanism. In addition, in Ref. [14\*\*] the authors also show that lack of amino acid sensing confers a negative fitness to *Drosophila melanogaster in vivo*.

- Harrington LS, Findlay GM, Gray A, Tolkacheva T, Wigfield S, Rebholz H, Barnett J, Leslie NR, Cheng S, Shepherd PR et al.: The TSC1-2 tumor suppressor controls insulin-PI3K signaling via regulation of IRS proteins. J Cell Biol 2004, 166:213-223.
- Hartley D, Cooper GM: Role of mTOR in the degradation of IRS-1: regulation of PP2A activity. J Cell Biochem 2002, 85:304-314.
- Haruta T, Uno T, Kawahara J, Takano A, Egawa K, Sharma PM, Olefsky JM, Kobayashi M: A rapamycin-sensitive pathway down-regulates insulin signaling via phosphorylation and proteasomal degradation of insulin receptor substrate-1. Mol Endocrinol 2000, 14:783-794.
- Takano A, Usui I, Haruta T, Kawahara J, Uno T, Iwata M, Kobayashi M: Mammalian target of rapamycin pathway regulates insulin signaling via subcellular redistribution of insulin receptor substrate 1 and integrates nutritional signals and metabolic signals of insulin. Mol Cell Biol 2001, 21:5050-5062.
- Um SH, Frigerio F, Watanabe M, Picard F, Joaquin M, Sticker M, Fumagalli S, Allegrini PR, Kozma SC, Auwerx J et al.: Absence of S6K1 protects against age- and diet-induced obesity while enhancing insulin sensitivity. Nature 2004, 431:200-205.
- Zhang H, Bajraszewski N, Wu E, Wang H, Moseman AP, Dabora SL, Griffin JD, Kwiatkowski DJ: PDGFRs are critical for PI3K/Akt activation and negatively regulated by mTOR. J Clin Invest 2007, 117:730-738.
- Carracedo A, Ma L, Teruya-Feldstein J, Rojo F, Salmena L, Alimonti A, Egia A, Sasaki AT, Thomas G, Kozma SC et al.: Inhibition of mTORC1 leads to MAPK pathway activation through a PI3K-dependent feedback loop in human cancer. J Clin Invest 2008, 118:3065-3074.
- Feng Z, Hu W, de Stanchina E, Teresky AK, Jin S, Lowe S, Levine AJ: The regulation of AMPK beta1, TSC2, and PTEN expression by p53: stress, cell and tissue specificity, and the role of these gene products in modulating the IGF-1-AKTmTOR pathways. Cancer Res 2007, 67:3043-3053.

- 24. Jones RG, Plas DR, Kubek S, Buzzai M, Mu J, Xu Y, Birnbaum MJ, Thompson CB: AMP-activated protein kinase induces a p53-dependent metabolic checkpoint. Mol Cell 2005, 18:283-293
- 25. Budanov AV, Karin M: p53 target genes sestrin1 and sestrin2 connect genotoxic stress and mTOR signaling. Cell 2008, **134**·451-460
- Freeman DJ, Li AG, Wei G, Li HH, Kertesz N, Lesche R, Whale AD, Martinez-Diaz H, Rozengurt N, Cardiff RD et al.: PTEN tumor suppressor regulates p53 protein levels and activity through phosphatase-dependent and -independent mechanisms. Cancer Cell 2003, 3:117-130.
- Manning BD, Cantley LC: AKT/PKB signaling: navigating downstream. Cell 2007, 129:1261-1274.
- Sarbassov DD, Guertin DA, Ali SM, Sabatini DM: Phosphorylation and regulation of Akt/PKB by the rictor-mTOR complex. Science 2005, 307:1098-1101.
- Huang J, Dibble CC, Matsuzaki M, Manning BD: The TSC1-TSC2 complex is required for proper activation of mTOR complex 2. Mol Cell Biol 2008, 28:4104-4115.

A regulatory role of TSC in mTORC2 activity is shown here. The authors describe that TSC positively regulates mTORC2 activity, hence, also indirectly activates Akt.

- 30. Dibble CC, Asara JM, Manning BD: Characterization of Rictor phosphorylation sites reveals direct regulation of mTOR complex 2 by S6K1. Mol Cell Biol 2009,
- 31. Choo AY, Yoon SO, Kim SG, Roux PP, Blenis J: Rapamycin differentially inhibits S6Ks and 4E-BP1 to mediate cell-type specific repression of mRNA translation. Proc Natl Acad Sci US A 2008. 105:17414-17419.
- 32. Feldman ME, Apsel B, Uotila A, Loewith R, Knight ZA, Ruggero D, Shokat KM: Active-site inhibitors of mTOR target rapamycinresistant outputs of mTORC1 and mTORC2. PLoS Biol 2009, 7:e38

See annotation to Ref. [35°].

 Garcia-Martinez JM, Moran J, Clarke RG, Gray A, Cosulich SC,
 Chresta CM, Alessi DR: Ku-0063794 is a specific inhibitor of the mammalian target of rapamycin (mTOR). Biochem J 2009. 421:29-42

See annotation to Ref. [35°].

- Thoreen CC, Kang SA, Chang JW, Liu Q, Zhang J, Gao Y,
- Reichling LJ, Sim T, Sabatini DM, Gray NS: An ATP-competitive mammalian target of rapamycin inhibitor reveals rapamycinresistant functions of mTORC1. J Biol Chem 2009, 284:8023-8032

See annotation to Ref. [35°].

Yu K, Toral-Barza L, Shi C, Zhang WG, Lucas J, Shor B, Kim J, Verheijen J, Curran K, Malwitz DJ et al.: Biochemical, cellular, and in vivo activity of novel ATP-competitive and selective inhibitors of the mammalian target of rapamycin. Cancer Res 2009, 69:6232-6240.

This reference, together with Refs. [32\*,33\*,34\*], describe the generation of clinically relevant small molecules that inhibit mTOR catalytic site. This work also shows an antitumoral effect of mTOR inhibitors in vivo, with apparently no toxicity in short-term assays.

- O'Reilly KE, Rojo F, She QB, Solit D, Mills GB, Smith D, Lane H, Hofmann F, Hicklin DJ, Ludwig DL et al.: mTOR inhibition induces upstream receptor tyrosine kinase signaling and activates Akt. Cancer Res 2006, 66:1500-1508.
- Ma L, Teruya-Feldstein J, Behrendt N, Chen Z, Noda T, Hino O, Cordon-Cardo C, Pandolfi PP: Genetic analysis of Pten and Tsc2 functional interactions in the mouse reveals asymmetrical haploinsufficiency in tumor suppression. Genes Dev 2005, 19:1779-1786.
- Manning BD, Logsdon MN, Lipovsky AI, Abbott D, Kwiatkowski DJ, Cantley LC: Feedback inhibition of Akt signaling limits the growth of tumors lacking Tsc2. Genes Dev 2005, 19:1773-1778.
- Skeen JE, Bhaskar PT, Chen CC, Chen WS, Peng XD, Nogueira V, Hahn-Windgassen A, Kiyokawa H, Hay N: Akt deficiency impairs

- normal cell proliferation and suppresses oncogenesis in a p53-independent and mTORC1-dependent manner. Cancer Cell 2006. 10:269-280.
- 40. Vasudevan KM, Barbie DA, Davies MA, Rabinovsky R, McNear CJ, Kim JJ, Hennessy BT, Tseng H, Pochanard P, Kim SY et al.: AKT-independent signaling downstream of oncogenic PIK3CA mutations in human cancer. Cancer Cell 2009. 16:21-32

Herein, support for the relevance of Akt-independent signaling downstream of PI3K in tumor cell lines and clinical samples is reported.

- Garcia-Martinez JM, Alessi DR: mTOR complex 2 (mTORC2) controls hydrophobic motif phosphorylation and activation of serum- and glucocorticoid-induced protein kinase 1 (SGK1). Biochem J 2008, 416:375-385.
- Sarbassov DD, Ali SM, Sengupta S, Sheen JH, Hsu PP, Bagley AF, Markhard AL, Sabatini DM: Prolonged rapamycin treatment inhibits mTORC2 assembly and Akt/PKB. Mol Cell 2006, 22:159-168.
- 43. Shor B, Zhang WG, Toral-Barza L, Lucas J, Abraham RT Gibbons JJ, Yu K: A new pharmacologic action of CCI-779 involves FKBP12-independent inhibition of mTOR kinase activity and profound repression of global protein synthesis. Cancer Res 2008, 68:2934-2943.
- Shiota C, Woo JT, Lindner J, Shelton KD, Magnuson MA: Multiallelic disruption of the rictor gene in mice reveals that mTOR complex 2 is essential for fetal growth and viability. Dev Cell 2006, 11:583-589.
- 45. Guertin DA, Stevens DM, Saitoh M, Kinkel S, Crosby K, Sheen JH, Mullholland DJ, Magnuson MA, Wu H, Sabatini DM: mTOR complex 2 is required for the development of prostate cancer induced by Pten loss in mice. Cancer Cell 2009, 15:148-159.

In this report, ablation of mTORC2 in mice, although impacting on tumor development, shows no obvious alterations in normal prostate gland, suggesting therapeutic mTORC2 inhibition could be well tolerated.

 46. Nardella C, Carracedo A, Alimonti A, Hobbs RM, Clohessy JG,
 Chen Z, Egia A, Fornari A, Fiorentino M, Loda M et al.: Differential requirement of mTOR in postmitotic tissues and tumorigenesis. Sci Signal 2009, 2:ra2.

This paper provides evidence for lack of toxicity by mTOR inhibition in normal adult cells in vivo, encouraging therapies based on mTOR inhibition

- 47. Guertin DA, Sabatini DM: The pharmacology of mTOR inhibition. Sci Signal 2009, 2:pe24.
- Brachmann S, Fritsch C, Maira SM, Garcia-Echeverria C: PI3K and mTOR inhibitors: a new generation of targeted anticancer agents. Curr Opin Cell Biol 2009, 21:194-198.
- Engelman JA, Chen L, Tan X, Crosby K, Guimaraes AR, Upadhyay R, Maira M, McNamara K, Perera SA, Song Y et al.: Effective use of PI3K and MEK inhibitors to treat mutant Kras G12D and PIK3CA H1047R murine lung cancers. Nat Med 2008,
- Ihle NT, Lemos R Jr, Wipf P, Yacoub A, Mitchell C, Siwak D, Mills GB, Dent P, Kirkpatrick DL, Powis G: Mutations in the phosphatidylinositol-3-kinase pathway predict for antitumor activity of the inhibitor PX-866 whereas oncogenic Ras is a dominant predictor for resistance. Cancer Res 2009, **69**:143-150.
- Torbett NE, Luna-Moran A, Knight ZA, Houk A, Moasser M, Weiss W, Shokat KM, Stokoe D: A chemical screen in diverse breast cancer cell lines reveals genetic enhancers and suppressors of sensitivity to PI3K isoform-selective inhibition. Biochem J 2008, 415:97-110.
- 52. Inoki K, Li Y, Xu T, Guan KL: Rheb GTPase is a direct target of TSC2 GAP activity and regulates mTOR signaling. Genes Dev 2003, **17**:1829-1834.
- 53. Inoki K, Zhu T, Guan KL: TSC2 mediates cellular energy response to control cell growth and survival. Cell 2003, **115**:577-590.
- Gwinn DM, Shackelford DB, Egan DF, Mihaylova MM, Mery A, Vasquez DS, Turk BE, Shaw RJ: AMPK phosphorylation of

- raptor mediates a metabolic checkpoint. *Mol Cell* 2008, **30**:214-226.
- Hemminki A, Markie D, Tomlinson I, Avizienyte E, Roth S, Loukola A, Bignell G, Warren W, Aminoff M, Hoglund P et al.: A serine/threonine kinase gene defective in Peutz-Jeghers syndrome. Nature 1998, 391:184-187.
- 56. Crino PB, Nathanson KL, Henske EP: **The tuberous sclerosis complex**. *N Engl J Med* 2006, **355**:1345-1356.
- Manning BD, Tee AR, Logsdon MN, Blenis J, Cantley LC: Identification of the tuberous sclerosis complex-2 tumor suppressor gene product tuberin as a target of the phosphoinositide 3-kinase/akt pathway. Mol Cell 2002, 10:151-162.
- Orloff MS, Eng C: Genetic and phenotypic heterogeneity in the PTEN hamartoma tumour syndrome. Oncogene 2008, 27:5387-5397.